

BIOL 240: General Microbiology

Spring 2020

March 23rd

<http://accounts.smccd.edu/staplesn/biol240/>

1. **QUIZ #5** due Friday night (first attempt).

2. **Midterm 2 will be returned THIS WEEK on CANVAS.**

1

Ch. 8: Microbial Genetics

OBJECTIVES: Students should be able to

1. **Ch. 8:** Describe the **Central Dogma** of molecular genetics and the **three processes** that drive the flow of genetic information in an organism.
 2. Describe several **properties of DNA** and the process of **DNA replication** that contribute to DNA's central role as the hereditary material.
 3. Draw a **replication fork** and label **5 enzymes involved in DNA replication**. Describe the function of each enzyme.
 4. Compare and contrast DNA Synthesis (**Replication**) with RNA Synthesis (**Transcription**). Why is it suitable that RNA is a less stable molecule than DNA? (**THINK: Structure → Function!!! ALWAYS!!**) [*Make a table!*]
 5. ** List the **three types of RNAs** and their functions in protein synthesis. How do they "read" the genetic code? Define a "**codon**", and explain the functional mechanisms of a **start codon** and a **stop codon**.
 6. ** Compare the stages of **Initiation, Elongation, and Termination** in **Replication, Transcription and Translation**. Indicate the direction of synthesis in each, and the names of the start and stop sites. [*Make a table!*]
 7. ** Diagram an **Operon**, including **6 DNA and protein components** important to its function. *Explain the purpose of this structure/organization.*
 8. Explain and give examples of **inducible & Repressible Operons**. *Describe what types of protein products or pathways each controls, and explain why this is appropriate.*
 9. List three types of possible **point mutations**, and explain how some types might be more severe than others. *Explain how can mutations be prevented or repaired.*
 10. Outline two methods for **identifying** and **isolating mutant** bacteria.
 11. Define, diagram & compare the **3 mechanisms of Horizontal Gene Transfer** in prokaryotes.
- ❖ **Objectives are your HOMEWORK between classes!! **Read, Review, Draw!!**
 > **Outline the concepts, define terms, DRAW structures and processes, and PRACTICE!!**

2

Chapter 8

Microbial Genetics

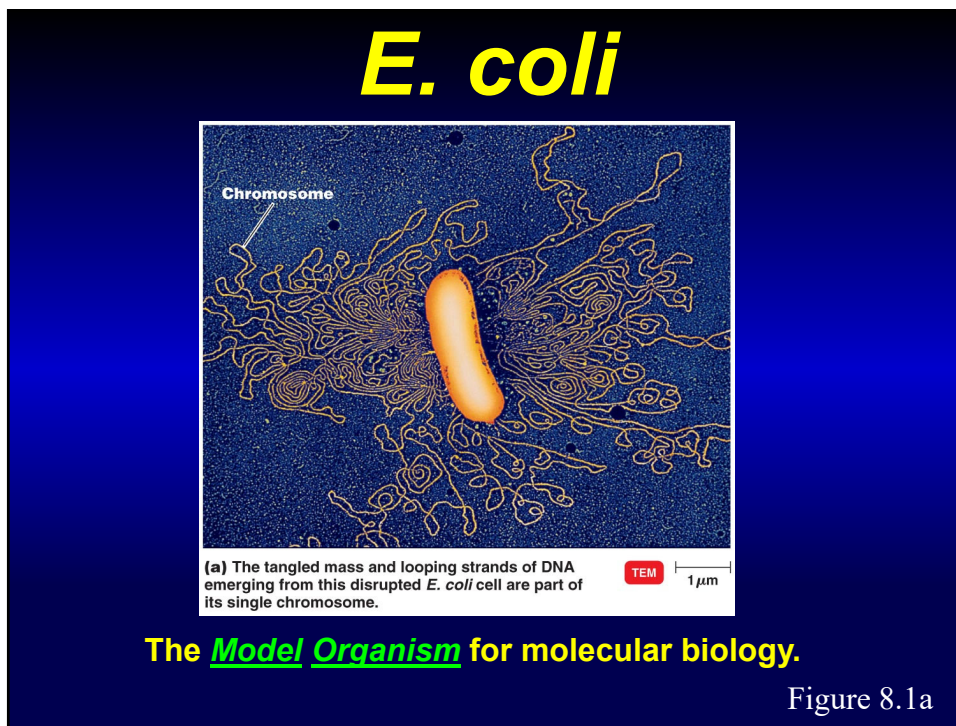


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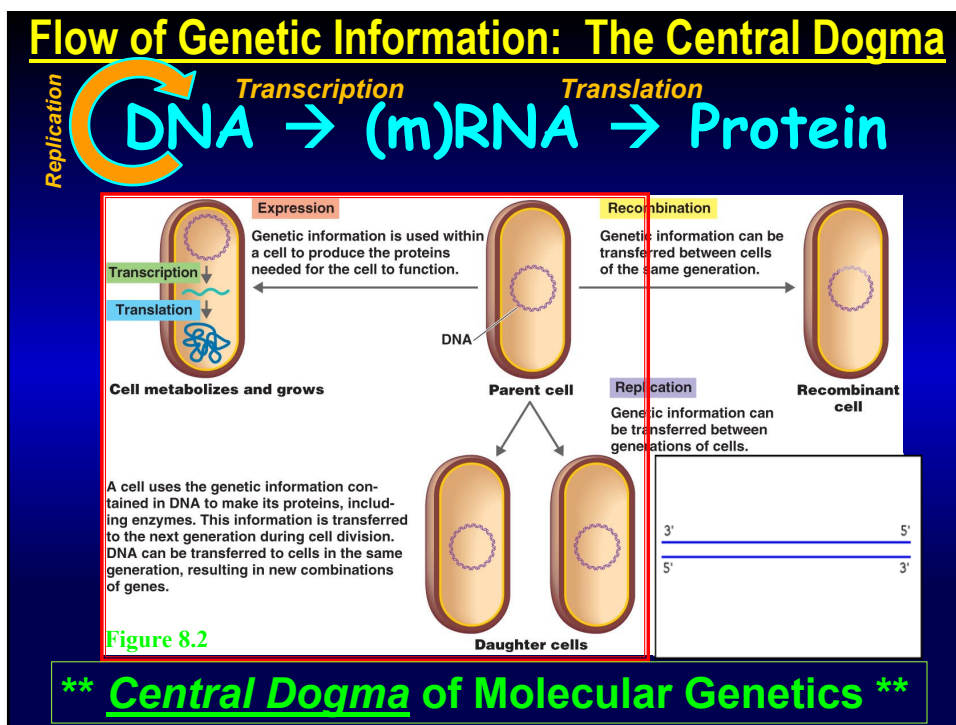
Terminology

1. **Genetics:** Study of what genes are, how they carry information, how information is expressed, and how genes are replicated; “the science of heredity”
2. **Gene:** Segment of DNA that encodes a functional product, usually a protein
3. **Genome** = All of the genetic material in a cell
4. **Genomics** = Molecular study of genomes
5. **Genotype** = Specific forms of genes in an organism
 - Types of alleles present.
6. **Phenotype** = *physical characteristics resulting from expression of the genes*

4



5

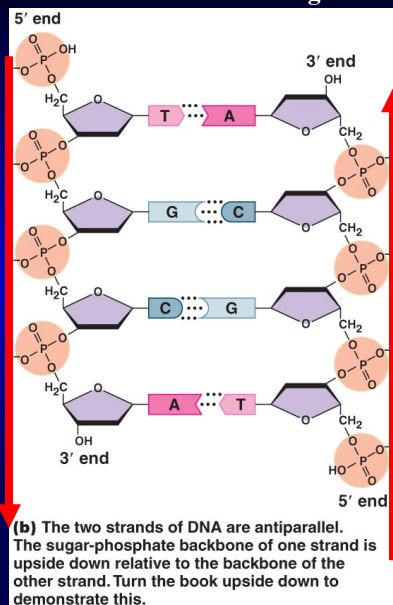


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8.1) DNA

1. Polymer of nucleotides: adenine, thymine, cytosine, guanine (**ATGC**)
2. Double helix associated with proteins
3. "Backbone" is deoxyribose-phosphate
4. Strands held together by hydrogen bonds between **A=T** and **G≡C**
5. Strands are **Antiparallel**
 - **5' (PO₄) → 3' (OH) polarity**

Figure 8.3

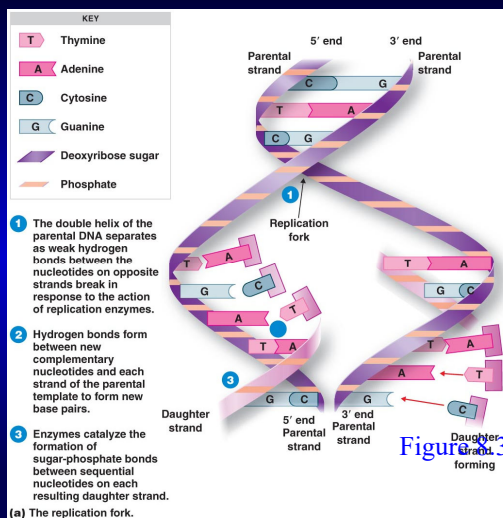


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A. DNA Structure

• **Semi-Conservative Replication:**

- Each “parental strand” serves as **template...**
- for synthesis of a new “daughter strand”
- Following **complementary base-pairing** rules
 - **A=T**
 - **G≡C**
- Rules allow one to predict 2nd strand sequence from 1st!!!



GREAT overview: <https://youtu.be/TNKWgcFPHqw>
http://www.uic.edu/classes/cmeng/cmeng521/DNA_replication.html

8

DNA Replication Proteins

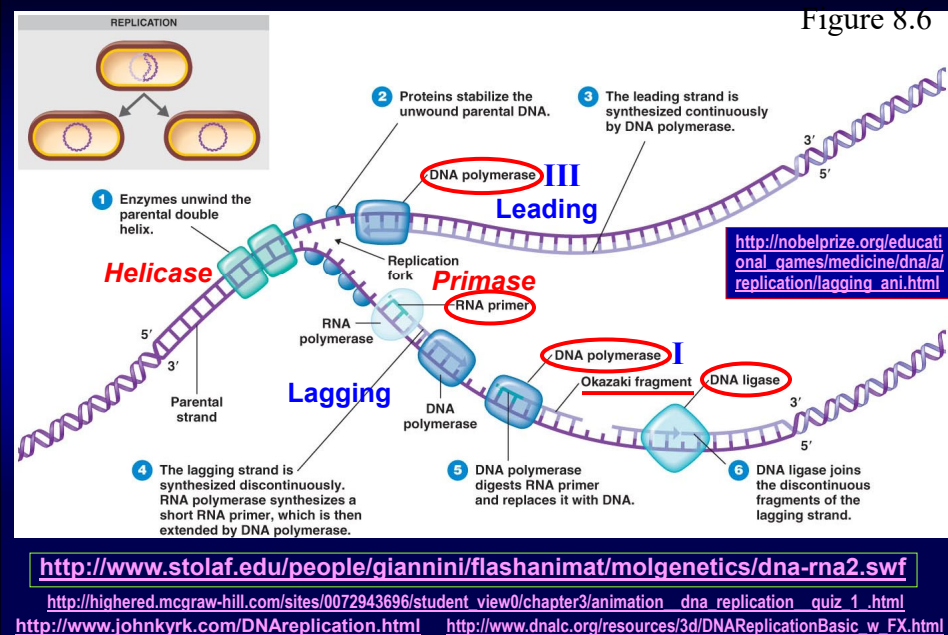
- Open helix at **Origin (Ori)**, lay-down primers:
 - DNA Helicase** "melts" strands apart, breaking H-bonds
 - Single-Strand Binding Proteins (**SSB**) keep template strands apart.
 - RNA Primase** lays down first several nucleotides (RNA!!) – gives "starting block" (free 3'-OH) to begin actual DNA synthesis. [RNA Primers are removed later!]
- DNA = copied by **DNA polymerase III (Dpol3)**
- In the **5' → 3'** direction – new nucleotides added to the 3' hydroxyl (-OH) group on deoxyribose in the growing strand – Initiated by an **RNA primer (RNA Primase enz.)**
- Leading strand** synthesized **continuously**
 - follows fork (1/fork; 2/ "bubble")
- Lagging strand** synthesized **discontinuously**
 - Opposite to fork movement → **Okazaki fragments (unsealed lagging pieces)**
- RNA primers are removed and Okazaki fragments joined by **DNA polymerase I & DNA ligase** → "fill and seal" to finish job!!

AMOEBASISTERS!!!!
<https://youtu.be/5qSrmeiWsuc>

- <http://www.stolaf.edu/people/giannini/flashanimat/molgenetics/dna-rna2.swf>
 Detailed: http://www.wiley.com/college/pratt/0471393878/student/animations/dna_replication/index.html

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C. DNA Replication Fork



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DNA replication is Semiconservative & Bidirectional

Figure 8.6
 (a) An *E. coli* chromosome in the process of replicating
 (b) Bidirectional replication of a circular bacterial DNA molecule

3D REALTIME:** <https://youtu.be/6j8CV3droDw>
<http://www.hhmi.org/biointeractive/dna/animations.html>

http://www.wornon.com/college/biology/mbio/animations/dna_replication.asp
<http://www.sciencemedia.com/website/demos/biochem/ecoli/Replication.html>

- = **DNA-directed DNA synthesis!!!**
- **2 forks: opposite directions!**
- Replication results in two daughter DNA duplexes,
 - each with one completely new strand, &
 - one old strand (parental strand)
 - = **“SEMI-CONSERVATIVE”**
- **ONLY synthesize 5' → 3'!!!**
- **Two replication forks** move in opposite directions =
 - ❖ **“Replication Bubble”**

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8.2) Transcription: RNA Synthesis

1. DNA is transcribed to make RNA (**AUGC**)
 - a) **mRNA** = messenger RNA → translated to protein
 - b) **tRNA** = transfer RNA → brings amino acid to ribos.
 - c) **rRNA** = ribosomal RNA → makes up ribo; catalytic
2. Transcription begins when **RNA polymerase** binds to the **PROMOTER** sequence
 - **Tells Rpol which strand, and where to start transcribing!!**
3. Transcription proceeds in the **5' → 3' direction** (same in ALL nucleic acid synthesis!)
 - new nucleotides added to the 3' hydroxyl group on **ribose** in the growing strand
4. Transcription stops when it reaches the **Terminator Sequence** (often many U's or “hairpin loop”)
 - New RNA and Rpol fall off of DNA template.

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<http://www.stolaf.edu/people/giannini/flash/animat/molgenetics/transcription.swf>

<http://vcell.ndsu.nodak.edu/animations/transcription/movie.htm>

TRANSCRIPTION

DNA
mRNA
Protein

RNA polymerase bound to DNA
AFM 6 nm

- 1 RNA polymerase binds to the promoter, and DNA unwinds at the beginning of a gene.
- 2 RNA is synthesized by complementary base pairing of free nucleotides with the nucleotide bases on the template strand of DNA.
- 3 The site of synthesis moves along DNA; DNA that has been transcribed rewinds.
- 4 Transcription reaches the terminator.
- 5 RNA and RNA polymerase are released and the DNA helix re-forms.

Promoter (gene begins)

RNA polymerase

Template strand of DNA

RNA

RNA-nucleotides

RNA synthesis

Terminator (gene ends)

Termination:
(eg.)

- polyU sequence, or
- Stem-loop in RNA

Figure 8.7

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8.3) Translation

1. = **RNA-directed Protein synthesis!!!**
2. mRNA is translated in **Codons.**
 - 3 nucleotide “words”
3. Translation of mRNA begins at the **start codon: AUG.**
 - Tells ribosome where to start translating,
 - and sets the reading Frame!!!
4. Translation ends at a **STOP (“nonsense”) codon: UAA, UAG, UGA.**
 - Do NOT encode an amino acid!!!

Transcription

Translation

Cell metabolizes and grows

Figure 8.2

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The "Universal" Genetic Code

Figure 8.8

		Second position				
		U	C	A	G	
U	UUU	Phe	UCU	UAU	UGU	Cys
	UUC		UCC	UAC	UGC	
	UUA	Leu	UCA	UAA Stop	UGA Stop	A
	UUG		UCG	UAG Stop	UGG Trp	
C	CUU	Leu	CCU	CAU	CGU	U
	CUC		CCC	CAC	CGC	
	CUA	Pro	CCA	CAA	CGA	A
	CUG		CCG	CAG	CGG	
A	AUU	Ile	ACU	AAU	AGU	Ser
	AUC		ACC	AAC	AGC	
	AUA	Met/start	ACA	AAA	AGA	Arg
	AUG		ACG	AAG	AGG	
G	GUU	Val	GCU	GAU	GGU	U
	GUC		GCC	GAC	GGC	
	GUA	Ala	GCA	GAA	GGA	A
	GUG		GCG	GAG	GGG	

- **Codons = non-overlapping**
 - Start transl'n; set the frame!!
- **tRNA (transfer RNA) =** adapter molecule that reads "3 letter" codons – via it's complementary "**Anticodon**"!
- Puts in the right amino acid in right order
 - **Reads mRNA 5' → 3'**,
 - **synthesizes N → C polypeptide**
 - **Amino → Carboxy!**
- **Anticodon-Codon base-pairing**

https://highered.mcgraw-hill.com/sites/0072507470/student_view0/chapter3/animation_how_translation_works.html
<http://www.lewport.wnyrc.org/jwanamaker/animations/Protein%20Synthesis.html>

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A. Translation – Initiation

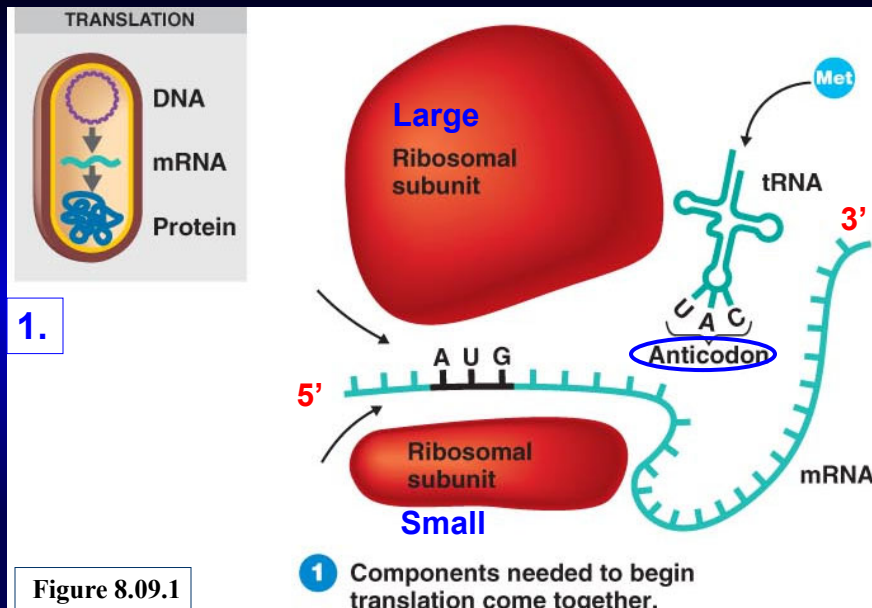


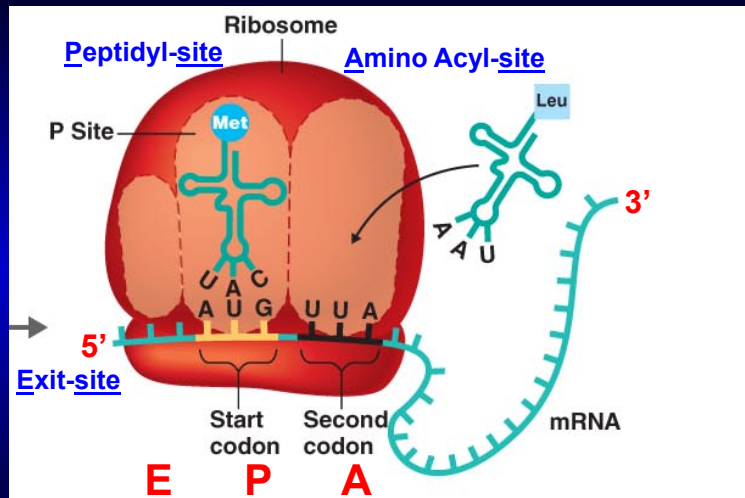
Figure 8.09.1

<http://www.stolaf.edu/people/giannini/flashanimat/molgenetics/translation.swf>
http://highered.mcgraw-hill.com/sites/0072437316/student_view0/chapter15/animations.html#

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B. Translation - Elongation

2.



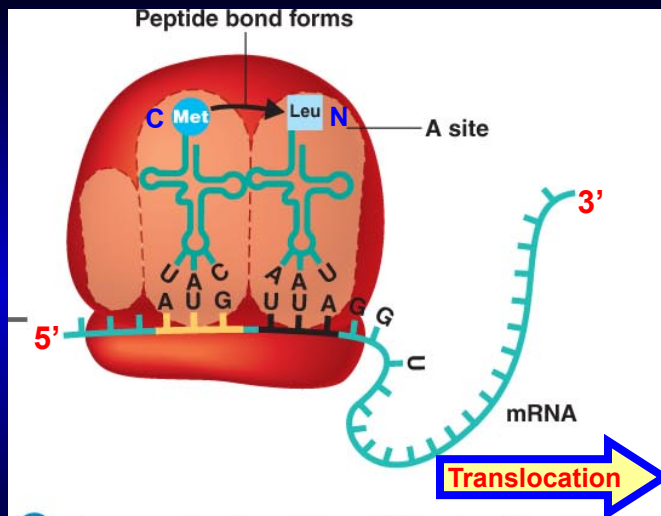
2 On the assembled ribosome, a tRNA carrying the first amino acid is paired with the start codon on the mRNA. The place where this first tRNA sits is called the P site.

Figure 8.09.2

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Peptidyl Transferase

3.



3 The second codon of the mRNA pairs with a tRNA carrying the second amino acid at the A site. The first amino acid joins to the second by a peptide bond. This attaches the polypeptide to the tRNA in the P site.

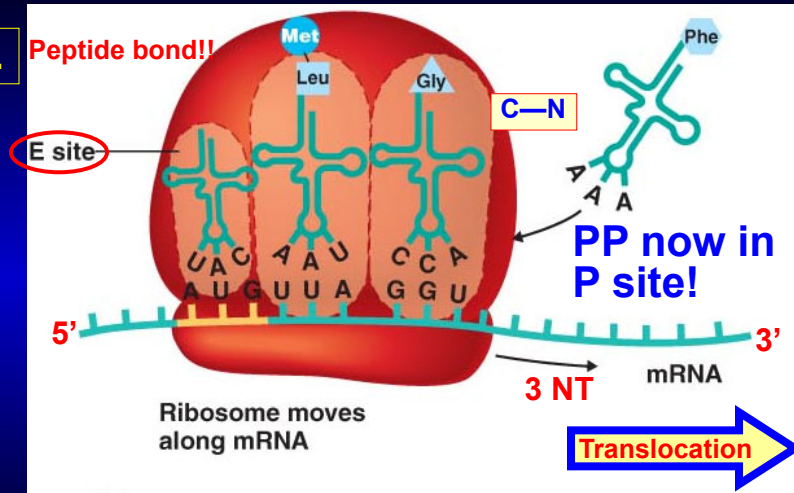
Figure 8.09.3

20

Newly elongated peptide in A site, then

4.

Peptide bond!!



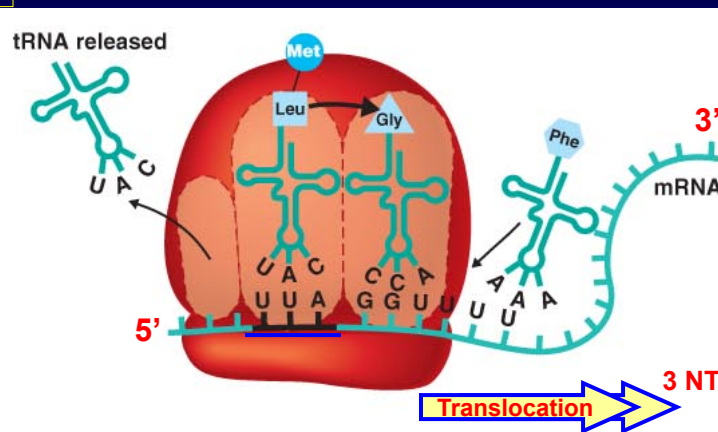
- 4 The ribosome moves along the mRNA until the second tRNA is in the P site. The next codon to be translated is brought into the A site. The first tRNA now occupies the E site.

Figure 8.09.4

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Ribosome Translocates 3nt in the 3' direction

5.



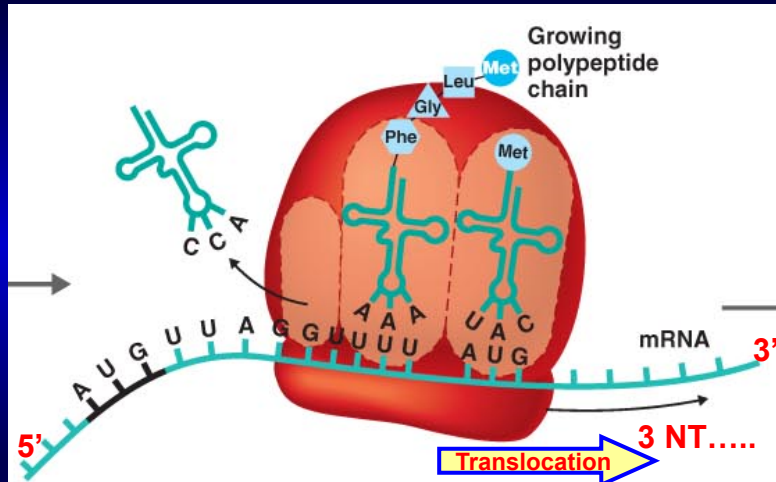
- 5 The second amino acid joins to the third by another peptide bond, and the first tRNA is released from the E site.

Figure 8.09.5

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Translation: Peptidyl Transfers & translocations continue

6.



6 The ribosome continues to move along the mRNA, and new amino acids are added to the polypeptide.

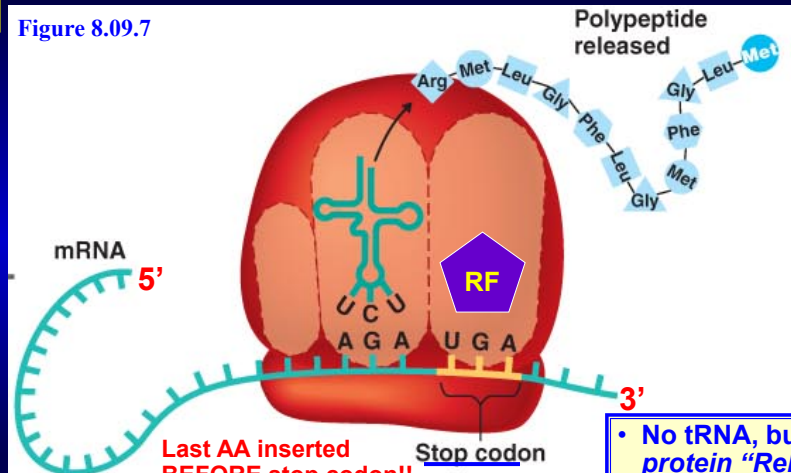
Figure 8.09.6

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C. Translation - Termination

7.

Figure 8.09.7



Last AA inserted BEFORE stop codon!!

Stop codon

7 When the ribosome reaches a stop codon, the polypeptide is released.

• No tRNA, but a protein "Release Factor" enters empty A site at a stop codon

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Tsln - Termination/ Release

8.

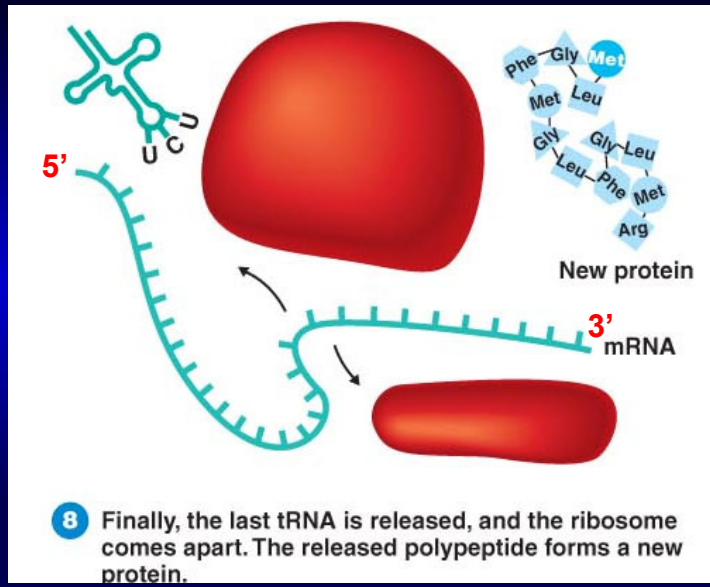
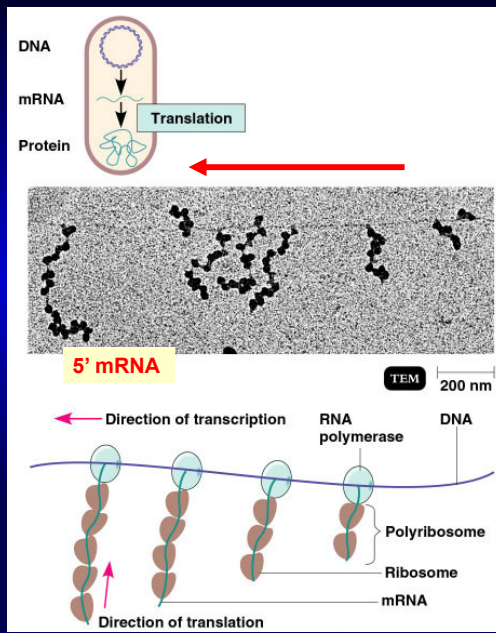


Figure 8.09.8

<http://vcell.ndsu.nodak.edu/animations/translation/movie.htm>

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** Polyribosomes



- Multiple ribos/ polypeptides translated per mRNA!!!
- = POLYRIBOSOMES (in both Euk. & Prok.!)

Also:

- Tsln & Tscn simultaneous in prokaryotes!! (only!)
- No nuclear membrane to separate!

Figure 8.10

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Compare Repln, Tscn, Tsln:

<u>Process</u>	<u>Initiation</u>	<u>Elongation</u>	<u>Termination</u>
<u>Replication</u>	At <u>Origin</u> : A/T-rich, NZs: Helicase, Primase, DPol3	Dpol3, dNTPs, 5'→3'	Terminator sequences, or end of chromosome (forks meet if circle)
<u>Transcription</u>	At <u>Promoter</u> : AT rich, NZ: RPol	RPol, NTPs, 5'→3'	Tsc'I terminator (poly-U, hairpin loop)
<u>Translation</u>	At <u>Start Codon</u> : (AUG), mRNA, AA-tRNA NZ: ribosome	Ribosome, AA-tRNA's (anticodons), N→C (follows mRNA 5'→3')	<u>Stop codon</u> (nonsense codon): UAA, UAG, UGA •Release Factor (= protein)

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8.4) Regulation of Bacterial Gene Expression

- Constitutive Genes (enzymes/proteins) are expressed at a fixed rate (“always on”!)
 - “housekeeping” functions in cells
- Other enzymes are expressed only as needed (“regulated enzymes/genes”)
 - Repressible Genes/enzymes = turned off when not needed; usually ON.
 - Inducible Genes/enzymes = turned on only when needed; usually OFF.

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Operons: Lactose Catabolism; Trp Synthesis



1 **Structure of the operon.** The operon consists of the promoter (P) and operator (O) sites and structural genes that code for the protein. The operon is regulated by the product of the regulatory gene (I).

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Figure 8.12.1

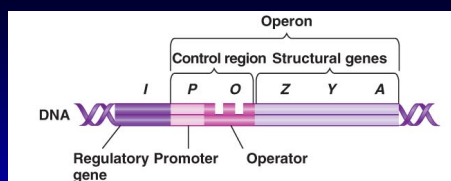
❖ **OPERATOR** = target/binding site on DNA.

❖ Bound by a **REPRESSOR** protein, to stop action of RNA Polymerase (→ genes OFF).

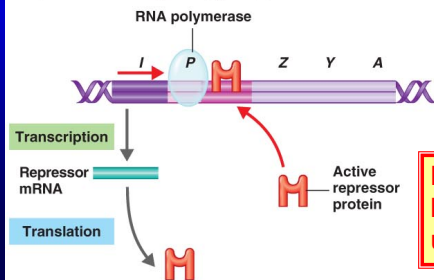
➤ = **Negative Regulation!**

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A. Lac Operon: Inducible



1 **Structure of the operon.** The operon consists of the promoter (P) and operator (O) sites and structural genes that code for the protein. The operon is regulated by the product of the regulatory gene (I).



Repressor protein usually bound to Operator (OFF!), unless lactose present.

2 **Repressor active, operon off.** The repressor protein binds with the operator, preventing transcription from the operon.

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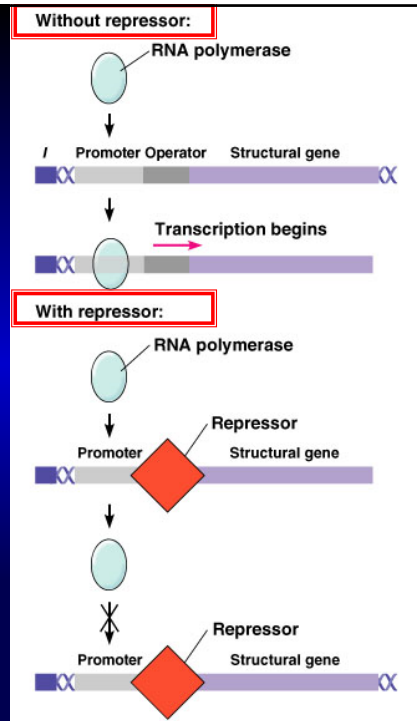
Figure 8.12.2

30

Repression

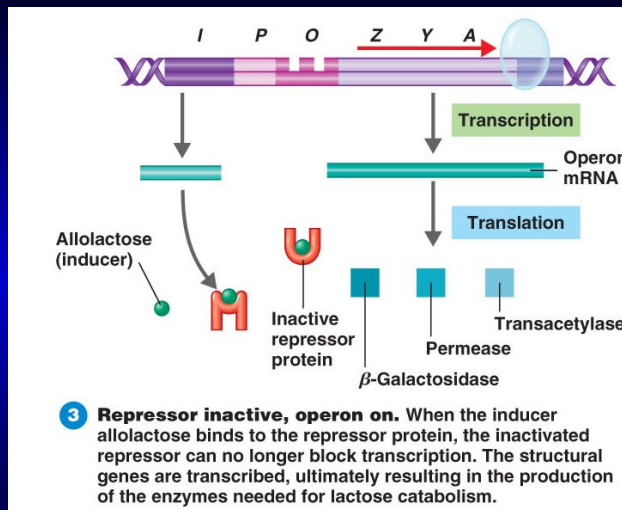
- Prokaryotes: Related genes organized into **OPERONS**:
 - Many “**structural genes**” (protein-encoding)
 - Under the control of a SINGLE **PROMOTER** & **operator** (negative control sequence)
 - **Polycistronic (polygenic) mRNA’s!**
 - Single mRNA transcript → encodes several proteins!!
 - NOT in EUKaryotes!!

Figure 8.13 (2004)



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Lac gene induction by Lactose



3 Repressor inactive, operon on. When the inducer allolactose binds to the repressor protein, the inactivated repressor can no longer block transcription. The structural genes are transcribed, ultimately resulting in the production of the enzymes needed for lactose catabolism.

Signal-Bound Repressor = OFF (genes ON), in an INDUCIBLE system

<http://vcell.ndsu.nodak.edu/animations/lacOperon/movie.htm>

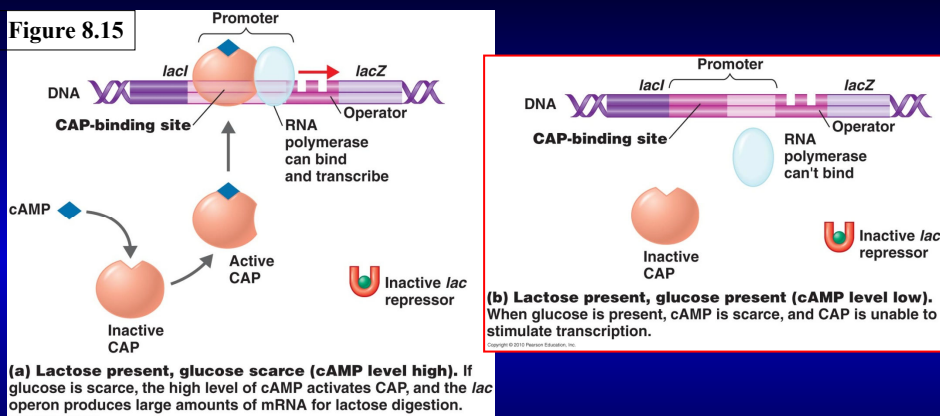
Figure 8.12.4

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Positive Regulation of the *lac* operon

http://www.biocourse.com/ui/swf/iLabs/lac_operon.swf***

Figure 8.15

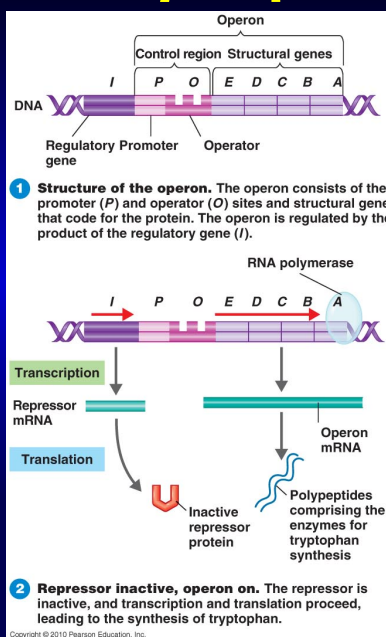


1. Glucose is the preferred energy source in most cells
2. For lactose and other alternative sugars to be consumed, glucose cannot be present
3. With glucose absent, ATP → ADP → AMP → cyclic AMP (starvation signal!!)
4. cAMP binds **CAP (Catabolite Activator Protein)**, which helps RPO_l bind *lac* promoter
5. **Full LAC operon activation requires Lactose PRESENT, and Glucose ABSENT!!!**

<http://highered.mcgraw-hill.com/olcweb/cgi/pluginpop.cgi?it=swf-535-535-/sites/dl/free/0072437316/120080/bio27.swf::Combination%20of%20Switches%20-%20the%20lac%20Operon>

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B. *Trp* Operon: Repressible



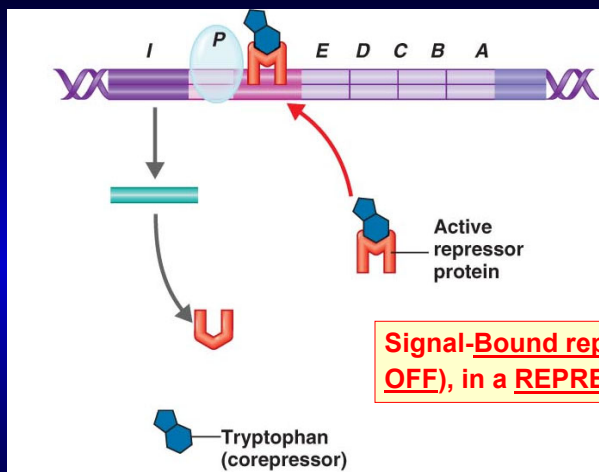
Repressor protein NOT bound to Operator, unless excess TRP present.

- **TRP = "corepressor"; activates repressor**
- → genes OFF when excess TRP

Figure 8.13

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Trp gene repression by excess TRP



<http://highered.mcgraw-hill.com/olcweb/cgi/pluginpop.cgi?it=swf::535::535::sites/dl/free/0072437316/120080/bio26.swf::The%20Tryptophan%20Repressor>

<http://bcs.whfreeman.com/thelifewire/content/chp13/1302002.html>

<http://nortonbooks.com/college/biology/animations/ch14a03.htm>

Signal-Bound repressor = ON (genes OFF), in a REPRESSIBLE system

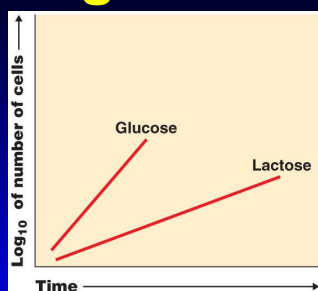
3 Repressor active, operon off. When the corepressor tryptophan binds to the repressor protein, the activated repressor binds with the operator, preventing transcription from the operon.

<http://highered.mcgraw-hill.com/sites/dl/free/0072835125/126997/animation28.html>

Figure 8.13

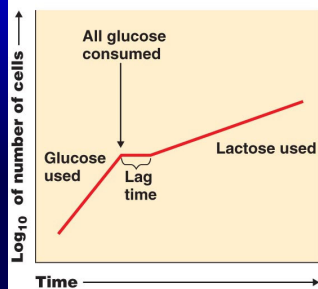
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Regulation of Gene Expression



(a) Bacteria growing on glucose as the sole carbon source grow faster than on lactose.

Faster growth on preferred C-source (glc)



(b) Bacteria growing in a medium containing glucose and lactose first consume the glucose and then, after a short lag time, the lactose. During the lag time, intracellular cAMP increases, the *lac* operon is transcribed, more lactose is transported into the cell, and β -galactosidase is synthesized to break down lactose.

Delayed growth (Lag!) on switching to secondary sugar, Lactose

- Lact catabolic enzymes must be induced & synthesized!!

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Operon Regulation: Summary

OPERON TYPE	ACTIVITY of	“Ligand” Molecule Absent	“Ligand” Molecule Present
Catabolic, ~lac <i>(Inducible)</i>	Repressor Protein	ON	OFF (lactose “inducer”)
	CAP Protein	OFF	ON (if ↑ cAMP; NO glucose!)
	Operon (transcription)	OFF	ON
Anabolic, ~trp <i>(Repressible)</i>	Repressor Protein	OFF	ON (tryptophan “corepressor”)
	Operon (transcription)	ON	OFF

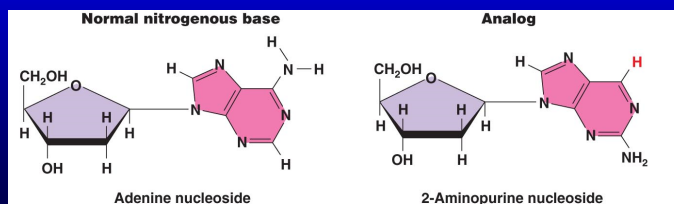
[Reminder: both signal molecules/ Ligands bind to the Repressor protein, **allosterically** changing its activity. Inducer, or Corepressor?]

- **Positive Control:** Regulatory Protein **ACTIVATES** genes.
- **Negative Control:** Regulatory protein **REPRESSES** genes.

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8.5) Mutation

- = **Change in the genetic material**
 - Source of evolutionary change!! (raw material)
 - The most powerful tool in Molecular Biology!!
- Mutations may be neutral, harmful, or beneficial.
- **Mutagen:** Agent that causes mutations above spontaneous levels.



(a) The 2-aminopurine is incorporated into DNA in place of adenine but can pair with cytosine, so an AT pair becomes a CG pair.

Fig. 8.19a

- **Spontaneous mutations:** Occur in the absence of a mutagen; due to normal, rare chemistry.

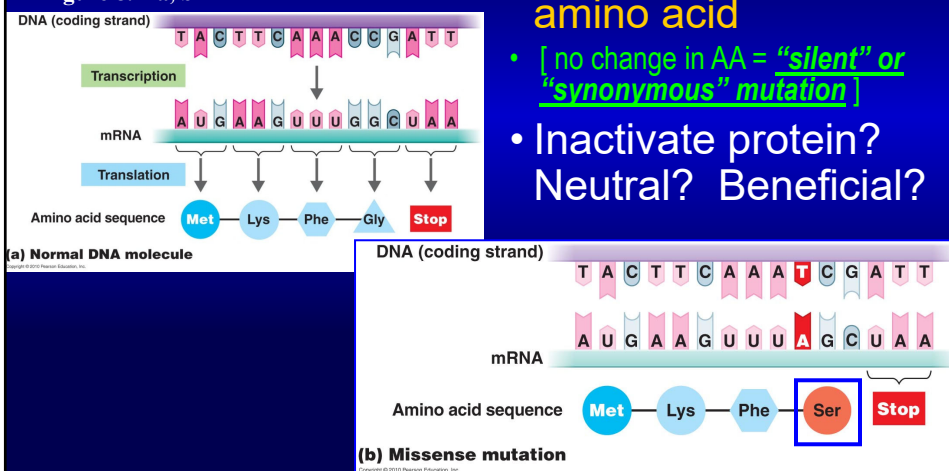
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A. Point Mutations (base substitution)

1. Missense mutation

- Change in one base
- Result in change in amino acid
- [no change in AA = "silent" or "synonymous" mutation]
- Inactivate protein? Neutral? Beneficial?

Figure 8.17a, b



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Mutation

2. Nonsense mutation:

- Results in a nonsense codon = STOP codon
- → Truncated protein!!
- = Inactive

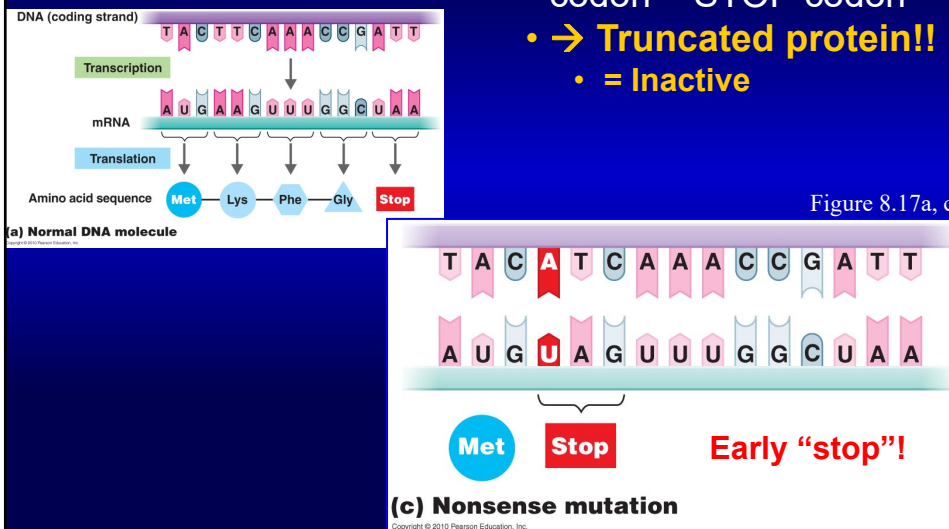


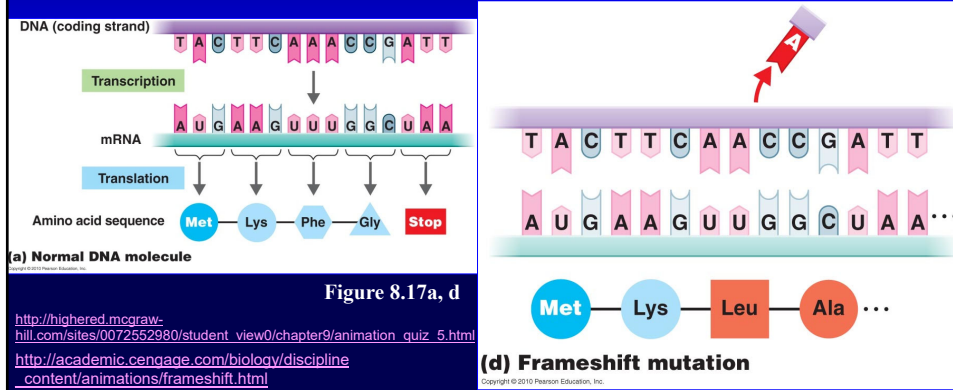
Figure 8.17a, c

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Mutation

3. Frameshift mutation:

- Insertion or deletion of one or more nucleotide pairs (when not multiple of 3) – severely change protein sequence from mutation site onward...



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Codons & Deletion Mutations

Normal

CAT CAT CAT CAT CAT CAT

• FAT CAT ATE THE RAT

- = DNA
- = Words analogy

One deletion

CTC ATC ATC ATC ATC AT

• FTC ATA TET HER AT.

Two deletions

CTA TCA TCA TCA TCA T

• FTA TAT ETH ERA T..

Junk!

Three deletions

CTA TAT CAT CAT CAT

• FTA TAE THE RAT ...

Back into Frame, -1 aa

3 deletions = mostly OK → clue codons = triplets!!

• Insertions or deletions (≠ multiple of 3), cause shift in reading frame

• = "Frameshift Mutation" → all codons downstream are JUNK!

• → inactive protein!

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B. Mutagenesis

- Ionizing radiation (X rays and gamma rays) causes the formation of ions
 - can react with nucleotides and the deoxyribose-phosphate backbone → DNA Damage!!
- **Nucleotide excision repair** fixes mutations

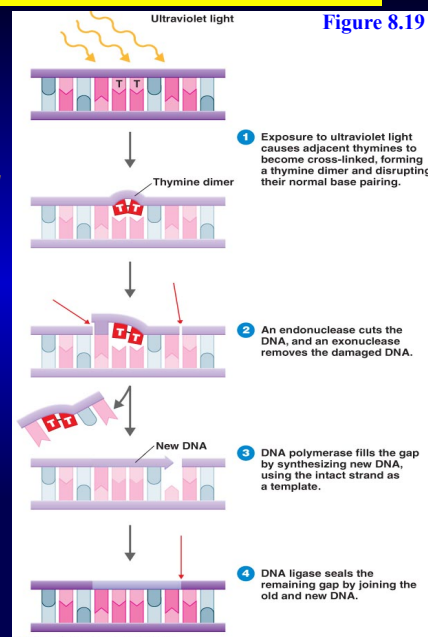
The Frequency of Mutation

- **Spontaneous mutation rate** =
 - 1 in 10^9 replicated base pairs (10^{-9}) or
 - ~1 in 10^6 replicated genes (10^{-6})
 - (assumes average gene = ~1000 bp)
- **Mutagens** increase to 10^{-5} or 10^{-3} per replicated gene (10-1000X more likely!).

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C.) DNA Excision Repair

- UV radiation causes **Thymine Dimers**
 - **Pyrimidine dimers!!!**
 - **Light-Activated Repair:** **Photolyases** separate thymine dimers.
1. **Endonuclease** enzyme clips DNA strand at each side of damage.
 - Recruited to site by **“Seeker Proteins”**.
 2. **Dpol1 & ligase** = fill & seal.



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D.) Studying Mutations: Selection

1. **Positive (direct) selection** detects mutant cells because they grow or appear different.

– eg: antibiotic-resistant mutants

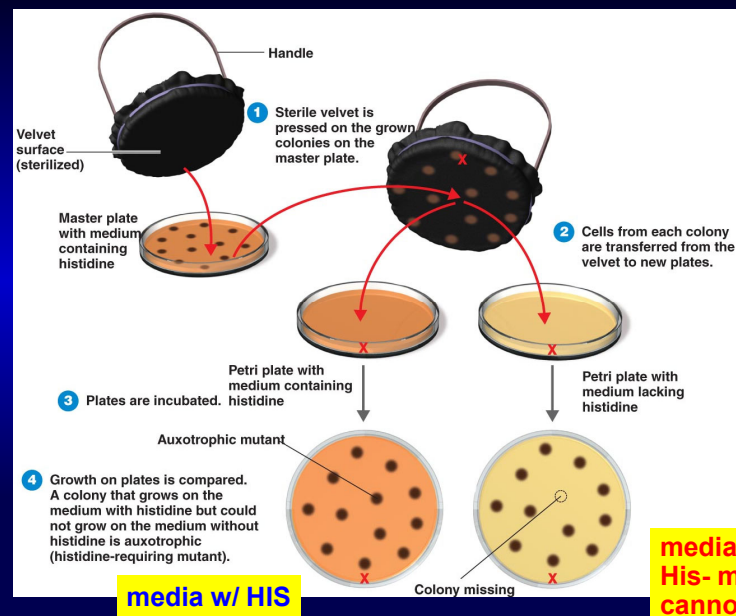
2. **Negative (indirect) selection** detects mutant cells because they do not grow.

– ** **Replica plating** – What does NOT grow on selective media?

- eg: **auxotrophic mutants** = need “help growing”, because lost ability to make a nutrient for themselves

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Replica Plating – for Neg. selection



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The Ames Test for Chemical Mutagens/Carcinogens

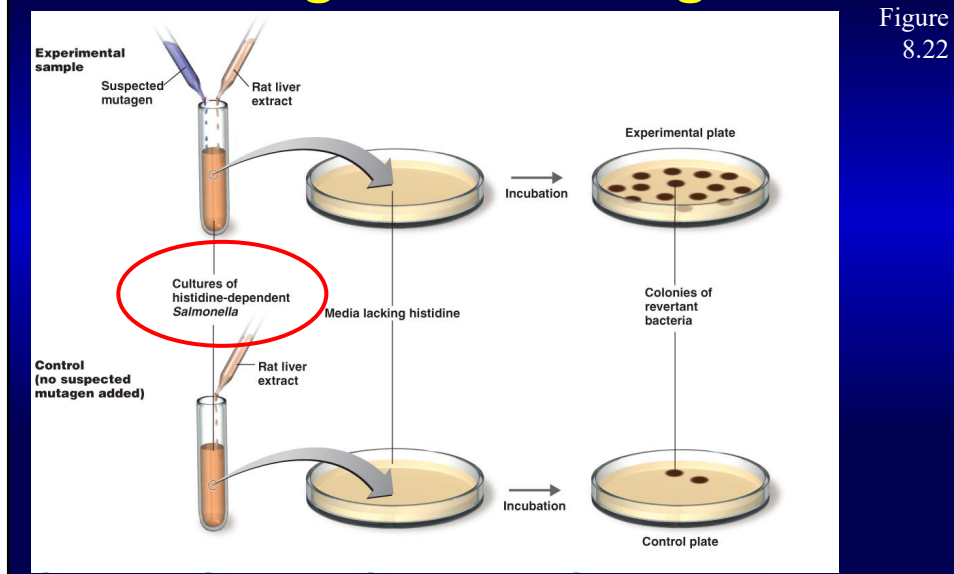
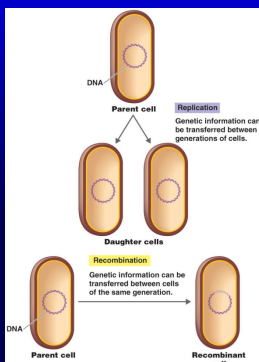


Figure 8.22

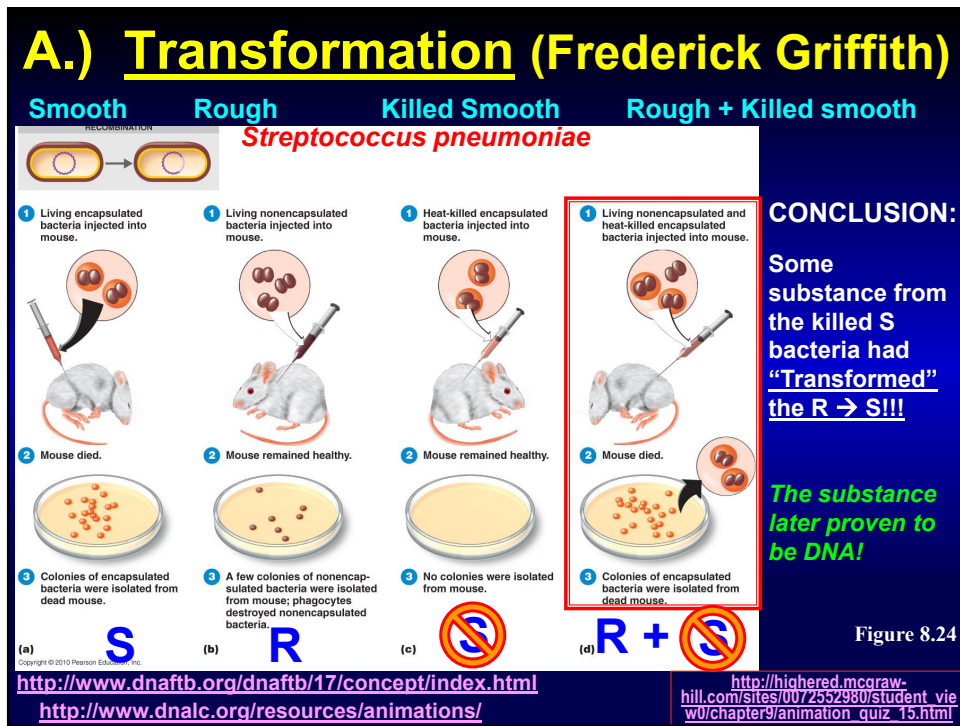
47

8.6) Genetic Transfer & Recombination

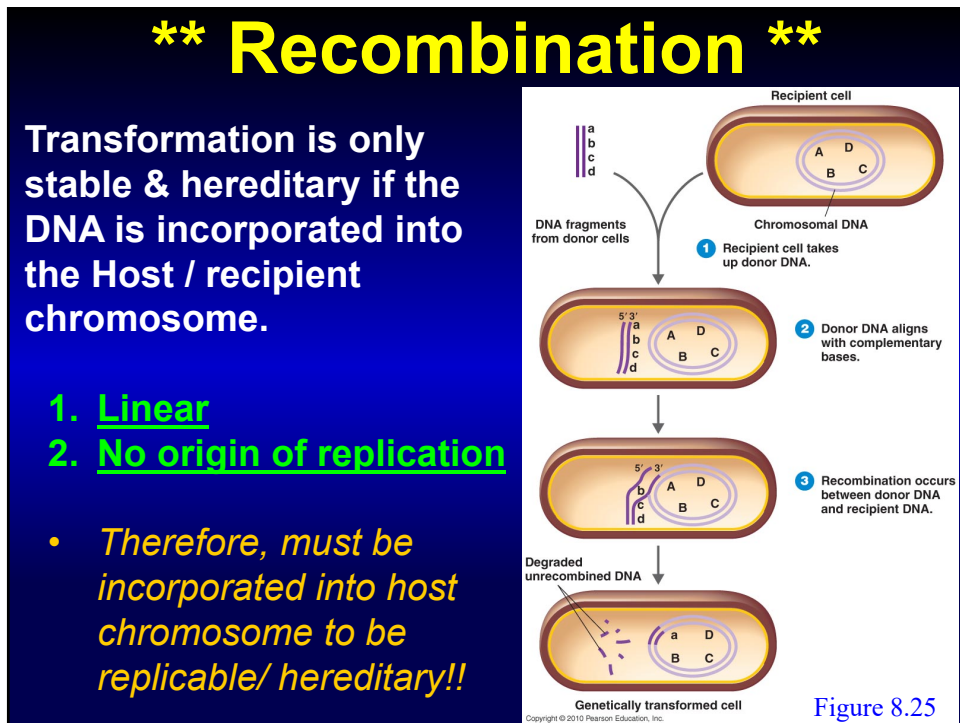
- **Vertical gene transfer** = Occurs during reproduction, between generations of cells.
- **Horizontal gene transfer** = Transfer of genes between cells of the same generation.



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** Genetic Recombination

- Exchange of genes between two DNA molecules
 - (Eg: *Meiosis-I in Eukaryotes*)
 - Crossing over occurs when two chromosomes break and rejoin
 - *** Bacteria & Archaea are Genetically Promiscuous cells!!!

- 1 DNA from one cell aligns with DNA in the recipient cell. Notice that there is a nick in the donor DNA.
- 2 DNA from the donor aligns with complementary base pairs in the recipient's chromosome. This can involve thousands of base pairs.
- 3 RecA protein catalyzes the joining of the two strands.
- 4 The result is that the recipient's chromosome contains new DNA. Complementary base pairs between the two strands will be resolved by DNA polymerase and ligase. The donor DNA will be destroyed. The recipient may now have one or more new genes.

Figure 8.23

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B.) Conjugation

Figure 8.27a

(Pilus = attachment only!!)

(transfer thru separate pore)

(a) When an F factor (a plasmid) is transferred from a donor (F⁺) to a recipient (F⁻), the F⁻ cell is converted to an F⁺ cell.

RECOMBINATION

- **F factor = Fertility Plasmid** (small, circular, self-replicating chromosome)
- Can carry genes from donor to recipient strains
 - F- recipient becomes F+

(a) Sex pilus

(b) Mating bridge

http://highered.mcgraw-hill.com/sites/0072556781/student_view0/chapter13/animation_quiz_3.html
Fig 8.26

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** Plasmids

1. Conjugative plasmid:

- Carries genes for sex pili and transfer of the plasmid

Mating of F+ and F-
Bacterial Strains

Animation by
Thomas M. Terry

2. Dissimilation plasmids:

- Encode enzymes for catabolism of unusual compounds
 - (*Pseudomonas* = toluene, camphor, petroleum HC's)

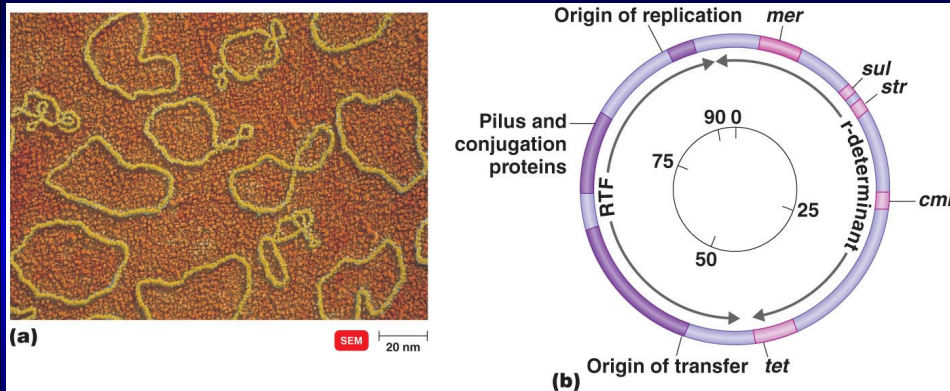
3. R factors:

- Encode antibiotic resistance (eg: MDR)

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Plasmids

Figure 8.29



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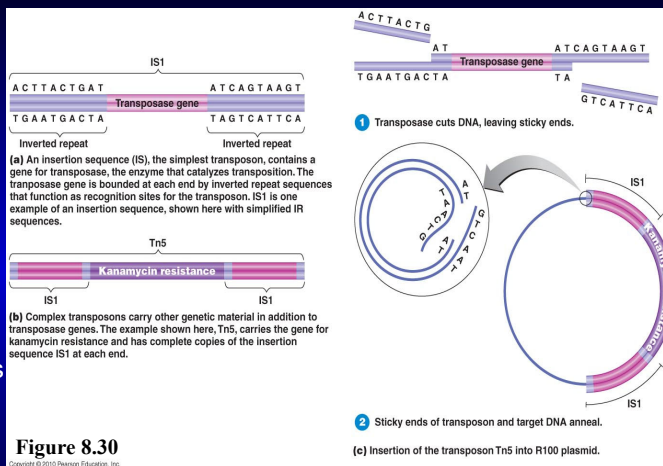
MDR Plasmid

<http://highered.mcgraw-hill.com/sites/dl/free/0072835125/126997/animation6.html>

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[Transposons]

- Segments of DNA that can move from one region of DNA to another
– “Jumping Genes”
- Contain **insertion sequences** for cutting and resealing DNA (**transposase**)
- Complex transposons carry other genes
– Eg: antibiotic-resistance



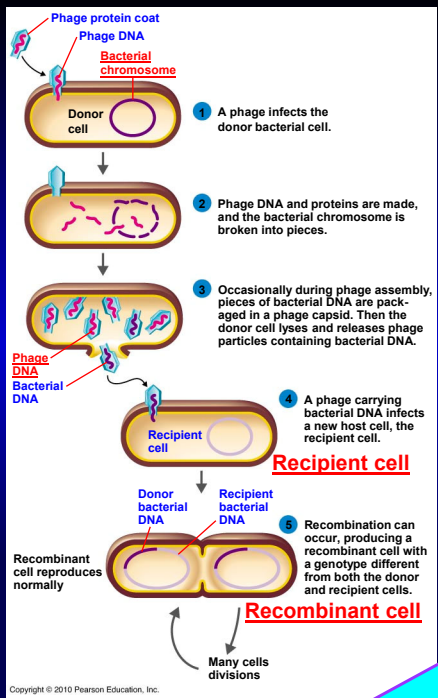
http://highered.mcgraw-hill.com/sites/0072556781/student_view0/chapter13/animation_quiz_5.html

<http://www.dnalc.org/resources/animations/alu.html>
<http://www.learner.org/courses/biology/archive/animations.html>

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C.) Transduction

Figure 8.28:
Transduction by a bacteriophage



<http://highered.mcgraw-hill.com/sites/dl/free/0072835125/126997/animation7.html>

http://highered.mcgraw-hill.com/sites/0072556781/student_view0/chapter13/animation_quiz_2.html

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